



## Letters

# Root-sprouting in myco-heterotrophic plants: prepackaged symbioses or overcoming meristem limitation?

Many plants spread by clonal growth. In most cases this is achieved using underground stems (e.g. rhizomes or stolons) which bear axillary buds at their nodes. New, potentially independent shoots (ramets) establish from these axillary buds. Clonal reproduction by the formation of adventitious shoots from roots is also possible (Groff & Kaplan, 1988) but is much less common. Only c. 10% of the species of the Central European flora are capable of forming adventitious buds on roots and sprout from them (Klimešová & Martíková, 2004), as particular hormonal and developmental barriers have to be overcome. Moreover, root sprouting plays different roles in the life cycle and growth form of individual plants (Wittrock, 1884; Rauh, 1937). In some, this is a necessary component towards completing their life cycle, for others it is an additional trait to 'normal' axillary stem branching, while in some this is a regenerative trait triggered by injury.

Necessary root sprouting may be found in very ecologically distinct groups; parasites, myco-heterotrophic plants and river weeds (Podostemaceae). In myco-heterotrophic plants, Domíniguez et al. (2006) discovered the formation of root propagules capable of adventitious sprouting, which soon detach from the parent plant and probably provide a clonal growth mode assuring the proper fungal symbiont to its vegetative offspring - 'prepackaged symbioses'. By clonal multiplication, a genet will produce more shoots and consequently more seeds leaving more sexual progeny, despite high seed and seedling mortality. Adventitious sprouting has been reported in several myco-heterotrophic species by several authors (Leake, 1994); however, the infection of propagules has not been studied and requires further examination. I wish to offer another explanation for root sprouting in myco-heterotrophs; this is not an alternative but rather a complementary idea to that of Domíniguez et al. (2006). The Central European flora will be used as an example. This flora contains a relatively small number of mycoheterotrophic plants, but the clonal growth modes of the plants are well known (Klimeš et al., 1997; Klimešová & Klimeš, 2006).

Dicotyledonous myco-heterotrophs and parasites in the Central European flora are characterised by a reduction of the primary stem in early ontogeny to such an extent that no axillary buds develop and plant growth is totally dependent on adventitious bud formation on the root or hypocotyle. Ecologically, the primary shoot is redundant in early ontogeny, as the plants draw carbon from hosts (parasites) or symbionts (myco-heterotrophic plants) below ground. The primary shoot may be lost in evolution in the case of overcoming meristem limitations by producing adventitious shoots later in ontogeny, merely to ensure sexual reproduction (full myco-heterotrophs; sensu Bidartondo, 2005) or for both sexual reproduction and carbon autonomy (initial mycoheterotrophs). For example, a myco-heterotrophic plant from the Ericaceae family, Monotropa hypopithys, germinates in the presence of a symbiont (Leake et al., 2004) with the embryo developing only a root pole. Roots supported by fungi form adventitious buds endogenously from the pericycle, close to the lateral roots (in stage 4 of germination according to Leake et al., 2004). Next-year buds sprout into adventitious shoots and, after flowering, die down to the parental root. Sprouting in the following year is again dependent on the formation of adventitious buds (Rauh, 1937). The plant perennates via the root system and adventitious sprouting results in clonal growth. No specialised root propagules are formed; however, new adventitious shoots arise from the infected root system. Therefore, no special means are needed to spread the symbiont into daughter plants as they remain connected to the parental root, which survives c. 3 yr (Beijerinck, 1887).

Another example is provided by representatives of the former Pyrolaceae family, which are only represented in Central Europe by photosynthetic plants (initial mycoheterotrophs). Moneses uniflora shows a similar ontogeny to Monotropa; however, its roots are thin and show a considerable lateral spread, while the aerial shoots survive several years of flowering (polycyclic shoots) (Irmisch, 1855; Rauh, 1937). Seedlings of Orthilia secunda germinate in a similar fashion to the preceding species: only by the root pole of the embryo (the same holds for all members of Ericaceae family listed in Table 1; Bobrov, 2004), and produce one adventitious shoot, which develops axillary buds for growth of a thin rhizome from which polycyclic shoots arise. Root sprouting from adventitious roots is also possible in the later stages of ontogeny (Irmisch, 1855); however, its contribution to clonal multiplication has never been quantified.

Unlike the dicotyledonous representatives of mycoheterotrophs in the Central European flora, orchids develop a primary shoot, form rhizomes on which there are axillary meristems for sprouting (Leake *et al.*, 2004) and, moreover, Table 1 List of mycoheterotrophic root-sprouters from the Central European flora with sources of morphological description

Species	Family	Source
Botrychium Iunaria (L.) SW.	Ophioglosaceae	13†
Botrychium virginianum (L.) SW.	Ophioglosaceae	13†
Ophioglossum vulgatum L.	Ophioglosaceae	3, 4, 12*, 15*
Cephalanthera alba (CRTZ.) SIMONKAI.	Orchidaceae	20
Cephalanthera longifolia (L.) FRITSCH	Orchidaceae	16
Cephalanthera rubra (L.) RICH.	Orchidaceae	12, 20*
Epipactis atrorubens (HOFFM.) BESSER	Orchidaceae	5†
Epipactis microphylla SWARTZ.	Orchidaceae	11†
Goodyera repens R. BR.	Orchidaceae	20†
Limodorum abortivum (L.) SW.	Orchidaceae	20*
Listera cordata (L.) R. BR.	Orchidaceae	12*, 17, 20*
Listera ovata (L.) R. BR.	Orchidaceae	17, 20*
Neottia nidus-avis (L.) RICH.	Orchidaceae	3, 12*, 20*
Spiranthes aestivalis RICH.	Orchidaceae	3†
Spiranthes autumnalis RICH.	Orchidaceae	3†
Monotropa hypopitys L.	Ericaceae	3*, 6, 9, 12*
Monotropa hypophegea WALLR.	Ericaceae	13†
Chimaphila umbellata (L.) BARTON	Ericaceae	7†
Moneses uniflora (L.) A.GRAY	Ericaceae	2, 6, 8*, 12*, 18, 19
Orthilia secunda (L.) HOUSE	Ericaceae	1, 6, 8*, 12*, 14*, 18
Pyrola chlorantha SW.	Ericaceae	8, 10

\*Picture available; +, only notice, without description.

<sup>1</sup>Bagdasarova (1990); <sup>2</sup>Bagdasarova & Vakhrameeva (1990); <sup>3</sup>Beijerinck (1887); <sup>4</sup>Filin (1995); <sup>5</sup>Summerhayes (1968); <sup>6</sup>Grevillius and Kirchner (1925); <sup>7</sup>Holm (1925); <sup>8</sup>Irmisch (1855); <sup>9</sup>Irmisch (1857); <sup>10</sup>Irmisch (1859); <sup>11</sup>Penzig (1921, 22); <sup>12</sup>Rauh (1937); <sup>13</sup>Jäger and Werner (2002); <sup>14</sup>Rysin and Rysina (1987); <sup>15</sup>Troll (1941); <sup>16</sup>Vakhrameeva *et al.* (1996); <sup>17</sup>Varligina (1995); <sup>18</sup>Velenovský (1905, 07); <sup>19</sup>Warming (1908); <sup>20</sup>Ziegenspeck (1936).

Species printed in bold are fully myco-heterotrophic, others are myco-heterotrophic only in early ontogeny. Other fully myco-heterotrophic plants from the flora (*Corallorhiza trifida* and *Epipogium aphyllum* from Orchideace) have no roots.

sprout adventitiously from roots (see sources in Table 1). They use two modes of clonal growth: while rhizomes are usually perennating for many years and do not spread far laterally, the roots are separated early from the parent plant and, in some species, they provide some lateral growth (Ziegenspeck, 1936; Filin, 1995). In these plants, adventitious sprouting is not necessary for completion of the plant life-cycle, but is an additive trait providing clonal multiplication, similar to the case described by Domíniguez *et al.* (2006). An interesting point, however, is that the orchids listed in Table 1 are the only root-sprouting monocots in Central Europe.

The coincidence of mycoheterotrophy and the ability to sprout adventitiously from roots is both ecologically and evolutionarily interesting and cannot be interpreted solely as an ecological adaptation. The role of the fungal symbiont in triggering root sprouting (the possible production of phytohormone-like substances; see Arshad & Frankenberger, 1991; Tsavkelova *et al.*, 2006), or their transport from a green host, has not been ruled out and needs to be taken into account, as it would provide a unifying explanation for the observed pattern. We can even speculate that root sprouting may be a by-product of the hormonal misbalance 'designed' for reversed carbon flow from the fungal symbiont to myco-heterotrophs. Adventitious bud formation is known to be triggered by a low auxin : cytokinin ratio (Banno *et al.*, 2001), which should be typical for myco-heterotrophs because auxin is produced by mature green and expanding leaves (Sachs, 2005).

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### References

Arshad M, Frankenberger WT. 1991. Microbial production of plant hormones. *Plant and Soil* 133: 1–8.

Forum

Bagdasarova TV, Vakhrameeva MG. 1990. Moneses uniflora. In: Vakhrameeva MG, ed. Biological Flora of Moscow Region 8: 181–188 [in Russian].

Banno H, Ikeda Y, Niu Q-W, Chua N-H. 2001. Overexpression of *Arabidopsis* ESR1 induces initiation of Shoot Regeneration. *Plant Cell* 13: 2609–2618.

Beijerinck MW. 1887. Wurzelknospen und Nebenwurzeln. Verhanelingen der Koninklijke Akademie Van Wetenschappen 25/3: 1–150.

Bidartondo MI. 2005. The evolutionary ecology of myco-heterotrophy. *New Phytologist* 167: 335–352.

Bobrov JA. 2004. On the early stages of ontogenesis of European Pyrolaceae species. *Botaniceskij Zhurnal* 89: 1342–1351 [in Russian].

Domíniguez L, Sérsic A, Melville L, Peterson RL. 2006. 'Prepackaged symbioses': propagules on roots of the myco-heterotrophic plant *Arachnitis uniflora. New Phytologist* 169: 191–198.

Filin VR. 1995. Ophioglossum vulgatum. In: Pavlov VN, Tikhomirov VN, eds. Biological Flora of Moscow Region 11, 4–36 [in Russian].

Grevillius AY, Kirchner O. 1925. Monotropaceae. In: Kirchner O, Loew E, Schröter C, eds. *Lebensgeschichte der Blütenpflanzen Mitteleuropas*. Eugen Ulmer Verlag, Band 4, Abt.1.181–243.

Groff PA, Kaplan DR. 1988. The relation of root systems to shoot systems in vascular plants. *Botanical Review* 54: 387–422.

Holm T. 1925. On the development of buds upon roots and leaves. Annals of Botany 39: 867–881.

Irmisch T. 1855. Bemerkungen über einige Pflanzen der deutschen Flora. *Flora* 13: 625–638.

Irmisch T. 1857. Ueber die Keimung und die Erneurungsweise von Convolvulus sepium und C. arvensis, so wie über hypokotylische Adventivknospen bei krautigen phanerogamen Pflanzen. Botanische Zeitung 15/28: 489–497.

Irmisch T. 1859. Kurze Mittheilungen über einige Pyrolaceen. *Flora* 32: 497–501.

Jäger EJ, Werner K. 2002. Exkursionsflora Von Deutschland. Heidelberg, Berlin: Spectrum Akademischer Verlag.

Klimeš L, Klimešová J, Hendriks R, van Groenendael J. 1997. Clonal plant architecture: a comparative analysis of form and function. In: de Kroon H, van Groenendael J, eds. *The Ecology and Evolution of Clonal Plants.* Leiden, the Netherlands: Backhuys Publishers, 1–29.

Klimešová J, Klimeš L. 2006. CLO-PLA3. http://clopla.butbn.cas.cz/.

Klimešová J, Martíková J. 2004. Intermediate growth forms as a model for

the study of plant clonality functioning: an example with root sprouters. *Evolutionary Ecology* **18**: 669–681.

Leake JR. 1994. The biology of myco-heterotrophic ('saprophytic') plants. New Phytologist 127: 171–216.

Leake JR, McKendrick SL, Bidartondo M, Read DJ. 2004. Symbiotic germination and development of the myco-heterotroph *Monotropa hypopitys* in nature and its requirement for locally distributed *Tricholoma* spp. *New Phytologist* 163: 405–423.

Penzig O. 1921–22. Pflanzen Teratologie, 2nd edn. Berlin, Zahlendorf: Verlag von Gebrüder Borntraeger.

Rauh W. 1937. Die Bildung von Hypocotyl- und Wurzelsprossen und ihre Bedeutung für die Wuchsformen der Pflanzen. Acta Nova Leopoldina, Halle 4/24: 395–555.

Rysin LP, Rysina GP. 1987. Morphology and Structure of Below-Ground Organs of Forest Herbs. Moskva: Nauka [in Russian].

Sachs T. 2005. Auxin's role as an example of the mechanisms of shoot/root relations. *Plant and Soil* 268: 13–19.

Summerhayes VS. 1968. Wild Orchids of Britain, 2nd edn. London: Collins.

Troll W. 1941–42. Vergleichende Morphologie der Höheren Pflanzen. Band 1 – Vegetationsorgane, Teil 3. Berlin, Zahlendorf: Verlag von Gebrüder Borntraeger.

Tsavkelova EA, Klimova SY, Cherdyntseva TA, Netrusov AI. 2006. Hormones and hormone-like substances of microorganisms: a review. *Applied Biochemistry and Microbiology* 42: 229–235.

Vakhrameeva MG, Varlygina TI, Kulikov PV. 1996. Cephalantera longifolia. In: Pavlov VN, Tikhomirov VN, eds. Biological Flora of the Moscow Region. 12: 48–59. [in Russian].

Varligina TI. 1995. Listera. In: Pavlov VN, Tikhomirov VN, eds. Biological Flora of the Moscow Region. 10: 52–63 [in Russian].

Warming E. 1908. The structure and biology of Arctic flowering plants I. Ericineae (Ericaceae, Pirolaceae). *Meddelelser Om Grönland* 36: 2–71.

Wittrock VB. 1884. Ueber Wurzelsprossen bei krautigarten Gewächsen, mit besonderer Rücksicht auf ihre verschiedene biologische Bedeutung. *Botanisches Centralblatt* 17(8,9): 227–232, 257–264.

Ziegenspeck H. 1936. Orchidaceae. In: Wangerin W, Schröter C, eds. Lebensgeschichte der Blütenpflanzen Mitteleuropas. Stuttgart: Eugen Ulmer, Version Band I, Abt. 4.

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Velenovský J. 1905–07. Vergleichende Morphologie der Pflanzen. Praha: Fr. Řivnáč Ver.